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Symptoms characteristic of epilepsy have been recognised relatively well at the very beginning of medicine. Despite that fact, mechanisms underlying the disease still remain unknown. The brain injury has been commonly considered as one of the most powerful triggers for epileptogenesis. It appeared, therefore, obvious that the application of neuroprotective drugs minimising effects of injury would also protect the brain from epileptogenesis. However, a question has been raised that the neuroprotective treatment might not always be beneficial to the injured brain, since it would also promote the survival of neurons of functional features permanently modified under pathological condition. Therefore, the neuronal survivors themselves might constitute the source of initial seizures or show an increased susceptibility to seizuregenic stimuli. Indeed, it has already been demonstrated that longterm effects of neuroprotective drugs might not always have a positive influence on further susceptibility of the injured brain to seizures. On the contrary, application of neuroprotectants during the course of epileptic seizures may instead result in smaller structural and functional impairments. Finally, effective stimulation of endogenous neuroprotective potential existing in the injured brain also became the subject of experimental verification. Apparently, physical exercise is a promising example of this therapeutic approach.

Key Words

Animal models of epilepsy; Neuroprotection; Brain injury

For many years, epilepsy has been a serious clinical problem. According to a WHO estimate, about 50 million people suffer from this disease worldwide. In recent years, there has been a rapid progress in its diagnosis and therapy due to the engagement of experts from various fields of pharmacology, surgery, neurology and neurobiology. New antiepileptic drugs, such as felbamate, gabapentin, lamotrignine, vigabatrin and many others1 were introduced thanks to the co-operation of interdisciplinary teams. Unfortunately, exact causes and mechanisms of epileptic discharges are still unknown, which greatly hinders attempts to efficiently counteract the development of this disease.

Animal models of epilepsy

Studies on epileptogenesis or new antiepileptic drugs require the use of animal models of epileptic seizures. In the search for an ideal model of epilepsy, more than 50 different methods of seizure induction have already been described. An ideal model should present a picture possibly the most closely resembling clinical symptoms in humans². Seizures should be accompanied by the changes in electroencephalographic activity of the brain. If a model fulfils these criteria, it can be used for studies of the mechanisms underlying epileptogenesis, and for the development of new treatment strategies. The pilocarpine model of temporal lobe epilepsy has been in use for about 25 years. This model was created by Waldemar Turski, a Polish scientist, who was the first to observe that the administration of the cholineraic agonist, pilocarpine, to rats induced limbic seizures manifested by an array of behavioural symptoms and characteristic changes in EEG. After acute phase of seizures, isolated seizures in animals which survived status epilepticus could even be observed for several years³. Histological analysis of brains of pilocarpine-treated animals demonstrated a number of pathological changes in the olfactory bulb, amygdala, thalamus, hippocampus and neocortex^{3,4}. An important weakness of the pilocarpine model is the high mortality of animals which can, however, be prevented by lowering pilocarpine doses and its combination with lithium chloride⁵.

Treatment with the glutamatergic agonist, kainic acid, is another method used to trigger epileptic seizures. Such treatment elicits several characteristic behavioural and electroencephalographic changes. Histological analysis shows lesions in brain regions similar to those caused by pilocarpine, but the changes are less conspicuous⁶.

Seizures can be provoked also by strychnine, pentetrazole, excitatory amino acids and many other substances⁵.

Recently, a research team led by Turski⁷ proposed a new model of epilepsy. Methomyl was used as a seizurogenic substance. It is a carbamate commonly used as an insecticide which can reversibly inhibit choline esterase activity. Methomyl in combination with lithium evokes limbic seizures followed by neurodegenerative changes and characteristic EEG alterations, but does not cause high mortality⁷. For this reason, this model can be an alternative to the models of epilepsy used so far.

Beside treatment with various substances, seizures can be provoked by a single electrical stimulation via ear clip electrodes⁸ or by repeated subthreshold stimulation (kindling). The latter method requires implantation of electrodes which *per se* injures the brain and may be epileptogenic.

The models of epileptic seizures presented above are most frequently used in experimental studies. Besides, there are a number of other models of epilepsy, e.g. models utilising visual or auditory stimulation, or genetic manipulation².

Post-traumatic epilepsy

Brain injury following ischemia or trauma is one

of the possible causes of epileptic seizures. Developmental malformations in the brain leading to permanent changes in its structure may also underlie epileptogenesis⁹.

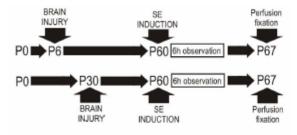
Clinical studies demonstrated that 50% of patients who suffered a penetrating brain injury developed epilepsy¹⁰⁻¹³. The risk of posttraumatic epilepsy increased with lesion severity, nevertheless, epilepsy could develop even in patients experiencing very small lesions¹⁴. Clinical studies, however, do not always guarantee effective exploration of effect of the injury on epileptogenesis since etiology of this disease in humans is usually not completely clear. Therefore, animal models of epilepsy or mechanical brain injury are used to elucidate this problem in more detail¹⁵⁻¹⁷. In these models, an effort is made to unambiguously define all parameters of seizures. Usually these studies are carried out on adult animals, therefore, it is difficult to speculate on that basis what effects such injury would elicit in the immature brain.

On the other hand, clinical studies report that children and teenagers are the groups in greatest danger of head injuries. The resulting brain injuries are the most frequent cause of neurological disorders (including posttraumatic epilepsy) in children^{17,18}.

For these reasons, recent studies of epileptogenesis focus on the search for means how to prevent development of epilepsy of different etiologies, particularly post-traumatic epilepsy in children and adults.

3. Experimental models of epilepsy 3.1. Pilocarpine model of epilepsy

In order to elucidate how mechanical injury influences brain susceptibility to epileptic seizures, a well-known model of penetrating injury of the brain hemisphere was used. In this model, the integrity of cranium and meninges was broken showing, therefore, a good analogy to sudden and traffic accidents¹⁹⁻²². Two animal groups were subjected to this kind of injury on postnatal days 6 or 30 (P6 and P30, respectively, Fig. 1 shows a general scheme of the experiment):





When the animals became 60 days old (P60s), they received pilocarpine injections to induce status epilepticus (SE)²³. Thereafter, all behavioural symptoms were assessed. It was proven that susceptibility to the seizures depended on the developmental stage at which the mechanical injury was performed.

In rats injured on P6, the course of seizures evoked on P60 did not differ from that observed in controls. On the contrary, brain injury on P30 severely increased susceptibility to seizures. Moreover, the proportion of survivors on the seventh day of the experiment in animals injured on P6, was higher than in the control group. On the contrary, mortality of animals injured on P30 was extremely high, and reached 97%²³. The evidence that injury on P6 did not increase the susceptibility to epileptic seizures evoked in mature animals does not agree with clinical data showing that paediatric insults are most epileptogenic^{12,24}.

Probably, diverse predisposition to seizures in animals injured on P6 or P30 is partially a result of different, age-dependent structure of the injury-induced glial scar. This is in agreement with pioneering research of Penfield²⁵, who first indicated the relation between astrogliosis and epilepsy. Maxwell and co-workers²⁶ proved that in animals injured in the first postnatal week, alial scar remains dispersed and permissive to neuronal regeneration process²⁷. Following the similar brain injury made in one-monthold animals, the glial scar has compact structure constituting a physical barrier for growing axons²⁸ making the restoration of damaged neural connections impossible. Apart from this, the scar is also a potent biochemical barrier producing axonal growth inhibitors²⁹. All the injury-induced structural and biochemical changes could be responsible for the significant differences in susceptibility to seizures between the two age groups. It is worth remembering that astrocytes have their own calciumdependent mechanisms of excitation, with surface receptors and ion channels, like in neuronal cells. For these reason, actrocytes can be considered as a primary source of epileptic activity³⁰. It may depend on the changes in proportion and functional interfaces between astrocytes and neurons, especially after injuryinduced glial proliferation and modification of extracellular environment.

Moreover, many cells within the region of injury undergo apoptotic or necrotic changes³¹. However, if they can survive, they may also be able to create additional pathological connections underlying increased susceptibility to spontaneous epileptic discharges or to seizuregenic stimuli and, therefore, facilitating occurrence of status epilepticus.

3.2. Kainic acid model of epilepsy

In clinical settings, we are not sure enough which neurotransmitter systems are involved in epileptogenesis and/or seizure processes. Animal models of seizures improve the situation by application of appropriate seizuregenic substances that can activate selected neuronal populations. Nevertheless, definition of specific neural subsystems responsible for changes in susceptibility to seizures can be much less effective. It is always necessary to keep in mind that the changes are observed in a given animal model following application of a specific seizure-generating substance that activates selected functional subsystems. Consequently, all the above-presented phenomena can rather be considered as characteristic of the pilocarpine model of epilepsy²³. In order to resolve, if the activation of another neurotransmitter systems can produce similar effects, another model of epileptic seizures using kainic acid was applied in the same experimental paradigm³². Since kainic acid is a glutamateraic agonist, its dosage was calculated on the basis of data from experiments by other authors, to produce similar histopathogical and behavioural changes^{6,33}. Contrary to previously described results obtained using pilocarpine model of epilepsy²³, adult rats with brains injured on P30 showed no change in their susceptibility to kainic acid-induced seizures. Furthermore, brain injury performed on P6 led to significant amelioration in the course of seizures in the same experimental conditions. The results indicate that the brain injury may have different influence on long-term susceptibility to seizures when agents used to evoke the seizures activate functionally different neuronal subsystems. Changes in susceptibility to seizures following brain injuries performed at different developmental stages may also present different profiles depending on the experimental model, i.e. when pilocarpine or kainic acid were chosen as seizuregenic agents^{23,32}. Each of the two models of epilepsy might display different aspects of the same age-dependent process triggered by the brain injury. These facts show clearly that no general and unequivocal conclusion concerning mechanisms of epipletogenesis can be obtained on the basis of experiments using a single experimental model of epilepsy since the process appears to depend on multiple functional subsystems.

Potential for use of CsA and FK506 in antiepileptic therapy

Since, as mentioned earlier, each brain injury may have irreversible consequences, factors that

could prevent development of post-traumatic changes have been tested for many years. Compounds that can minimise the effects of brain injury by increasing the ability of nervous cells to survive under pathological conditions are defined as neuroprotective factors.

In experimental studies, such substances can either be administered systemically or the organism itself can be stimulated to produce neuroprotective factors. If such factors are considered to be used in clinical practice, they should be thoroughly examined in preclinical studies. Unfortunately, only a few of them fulfil requirements for being used to treat humans.

Immunosuppressive action of CsA and FK506

CsA was first described as neuroprotective compound based on experimental studies by Shiga et al³⁴. Until then, it had been exclusively used as an immunosuppressant. CsA is a lipophilic undecapeptide isolated from the fungus *Tolrypocladium infantum* Gams in 1976³⁵. It has a wide-ranging biological action as a fungicide, and an antiparasitic or anti-inflammatory agent. Finally, it is used as an immunosuppressant in transplantology^{36,37}, in autoimmune diseases of the skin³⁸ and of the nervous system³⁹ and in rheumatism⁴⁰.

FK506 (Tacrolimus, commercial name Prograf), isolated from the fungus *Streptomyces tsukubensis* in 1984, shows similar actions to CsA⁴¹. It has been successfully used for the prevention of liver and heart graft rejection. When this drug is used, it is not necessary to administer so high doses of corticosteroids as in posttransplantational therapy with CsA⁴².

The main therapeutic effect of the two compounds consists in the inhibition of lymphocyte I activation^{43,44}. When CsA or FK506 have entered the cell, they bind to cyclophilins (CyPs) or FK binding proteins (FKBPs), known also as immunophilins⁴⁵. CyPs or FKBPs complexes are able to inhibit the activity of calcineurin, which is an enzymatic protein with phosphatase activity. Because of its ability to cleave phosphate groups, it is engaged in many metabolic reactions critical for cell function.

Calcineurin is involved in the regulation of ion channel activity⁴⁶, and controls neurotransmitter release⁴⁷ and nitric oxide synthase activity⁴⁸, moreover, it is engaged in signal transmission in the cell⁴⁹.

The activated calcineurin dephosphorylates nuclear factor of activated T cells (NFAT), among other molecules, which leads to conformational changes exposing site of NFAT transport to the nucleus. NFAT has relatively low affinity for DNA and interacts with other transcription factors to activate such promoters as IL-2,IL-3, GM-CSM, TNF and Fas ligand, which initiate T cell activation⁵⁰. Therefore, if calcineurin is inhibited, IL-2 cannot be activated, which disables full activation of T cells^{51,52}. These phenomena are the basis of wellestablished immunosuppressant action of CsA and FK506.

By inhibiting calcineurin activity, CsA and FK506 affect also the activity of another transcription factor CREB (cAMP response element-binding), which amplifies their immunosuppressant activ-ity^{53,54}.

4.2. Neuroprotective action of immunosuppressive compounds

Many years of the studies of neuroprotective action of the above-mentioned immunosuppressive compounds paved the way for their use in different models of brain (traumatic brain injury, global and partial ischemia) or peripheral nervous system injury.

4.2.1. Neuroprotectine action of cyclosporine

The experiments presented below illustrate the action of neuroprotective compounds in the selected models of brain injury that most closely simulate clinical situations.

CsA (10 mg/kg) administration for seven days before ischemia induction significantly reduced infarct area. Most probably, it was attributable to the activation of TrKa receptor of brain-derived neurotrophic factor (BDNF)⁵⁵. CsA injection significantly increased the level of phosphorylated CREB (pCREB), which was earlier observed in hippocampal cells resistant to hypoxia-induced damage^{56,57}. CsA at 20 mg/ kg decreased number of destroyed striatal dopaminergic neurons in an animal model of Parkinson's disease⁵⁸. CsA (10 mg/kg) administered after hypoxia induced by middle carotid artery occlusion (MCAO) significantly reduced degeneration area⁵⁹.

In the peripheral nervous system, CsA (2.5 mg/kg) administered 72 h before injury and an hour afterwards decreased demyelisation and number of vanishing nervous cells and supported recovery of motor functions⁶⁰. It also lowered proliferative activity of astrocytes in the injured brain (our unpublished studies).

4.2.2. Neuroprotective action of FK506

FK506 (1 mg/kg) administered after sciatic nerve section facilitated regeneration of pe-

ripheral axons by increasing the level of growth factors⁶¹. Likewise, a single injection of FK506 (3 mg/kg) 30 min before traumatic brain injury significantly diminished number of damaged neurons⁶². It reduced the area of hypoxiainduced damage in MCAO model even by 58% but only when it was administered in the dose range from 1 mg/kg to 10 mg/kg and not later than 2h after hypoxia63,64. The latest experiments have indicated that the mechanism of neuroprotective action of immunosuppressive drugs can depend on non-neuronal components of the nervous tissue. Specifically, it was noted that post-injury FK506 treatment modulated activity of astrocytes and microglia which was accompanied by lowering of astrocytic IL-1b mRNA and TNFa mRNA levels65. Moreover, astrocytic proliferation was reduced in the brains of animals that were treated with FK506 after mechanical brain injury (our unpublished results). It is probable that a combined treatment with FK506 and tissue plasminogen factor can be helpful in clinical treatment of brain ischemia⁶⁶.

A few examples of studies of neuroprotective action of CsA and FK506, presented above, provide evidence that these two compounds show neuroprotective activity in adult animals only when they are applied at appropriate time after injury. The time elapsing between the injury and treatment should be short enough to allow for onset of neuroprotective agent's action before irreversible changes actually occur (therapeutic time window⁶⁷). Obviously, the neuroprotective drugs should also be administered at appropriate doses that assure the most beneficial therapeutic effects with tolerable negative side effects for the organism.

Effect of neuroprotective compounds on epilepsy progression

5.1. Posttraumatic neuroprotection

Experimental studies repeatedly demonstrated neuroprotective action of CsA and FK506. Administration of these compounds after brain injury, which minimizes injury consequences, i.e. decreases necrosis area, reduces number of vanishing neurons, inhibits development of glial scars, could be the basis for more advanced clinical therapy.

Taking into consideration epileptogenicity of brain injury, known from clinical practice¹³, and varying post-injury vulnerability to epileptic seizures in animals²³, we decided to check whether the administration of neuroprotective compounds after brain injury would change predisposition to manifestation of seizure activity. To attain this goal, animals, whose brains were injured at different periods of postnatal life (P6 or P30), were administered cyclosporine A or FK506. Subsequently, when the animals were 60 days old, they were administered pilocarpine to induce seizures. We observed that pilocarpine-evoked seizure activity in 60-dayold animals with brains injured on P6 administered FK506 did not significantly differ from those which did not receive any neuroprotectant after injury. However, animals injured on P6 but treated with CsA after injury, showed aggravated seizures elicited on postnatal day 60 and mortality in this group was extremely high. On the other hand, post-injury FK506 treatment following brain injury performed on P30 did not significantly affect seizures evoked in adulthood but survival rate was increased. CsA treatment after injury in this age group prolonged seizure activity but markedly increased survival time after experimentally induced epilepsy⁶⁸.

These results did not provide full confirmation of the initial hypothesis assuming lighter seizures after treatment with neuroprotectants, but they proved that CsA and FK506 induced persistent changes in the damaged nervous tissue thereby changing its functional properties. It was manifested by the increased susceptibility to seizures, particularly after CsA treatment, whereas FK506 injection did not cause such changes but significantly prolonged survival after acute phase of seizures⁶⁸.

The question arises, therefore, whether final effects of neuroprotective intervention, evaluated only quantitatively (larger number of neurons protected from degeneration) is always functionally beneficial, since this is functional but not anatomical (structural) improvement which would justify implementation of neuroprotective strategy. Results of experimental studies usually are considered to be quite satisfactory if the number of surviving neurons in some damaged areas of the brain is increased or necrotic area is reduced. However, the studies presented above showed that the postinjury administration of compounds with welldefined neuroprotective action could have increased brain susceptibility to seizures. Therefore, it is possible that neuroprotection, whose efficacy is conspicuous at quantitative level and demonstrable by histological studies, not always leads to functional improvement of damaged brain, but, surprisingly, can even cause functional deterioration, since characteristics of 'rescued' neurons had been permanently modified by pathological posttraumatic conditions.

It should be remembered that cyclosporin A

and tacrolimus have non-polar molecules and thus they are not soluble in water. In posttransplational therapy, these drugs are administered in appropriate pharmaceutical formulations to achieve therapeutic effect⁶⁹. Substance in which an active substance is dissolved or suspended is called vehicle. In this case, a mixture of NaCl, Cremofor and ethyl alcohol was used as vehicle. Cremofor (modified castor oil) is the main component of the vehicle. Cremofor is obtained from Ricinus communis seeds and is characterised by very high viscosity and contains glycerides of ricinoleic acid, stearates and dihydroxystearic acid⁷⁰. Such vehicle composition increases stability of the drug stored in the form of solution. Moreover, Cremofor is a surface-active agent increasing solubility of the active compounds⁷¹. The vehicle contains also 0.25% ethanol in 0.9% NaCl. It improves drug solubility and uptake⁷².

It seems obvious that in order to be able to unequivocally assess CsA and FK506 potentials as the agents preventing seizure development after brain injury, it is crucial to test biological effects of vehicle in the same experimental design.

5.2. Effect of CsA and FK506 on acute phase of epilepsy

Clinical studies and animal models of epilepsy indicate that even a short episode of epileptic convulsions can produce local or remote neurodegenerative changes⁵. Neuronal death in vulnerable brain structures in rats can be observed already after epileptic discharges (in EEG recording) of 4-16 min duration^{73,74}. Seizure activity probably leads to the activation of postsynaptic NMDA receptors due to excessive glutamate release from presynaptic structures, and excessive calcium influx inside neurons, which can elicit numerous transformations, even cell death.

Hence, minimisation of the changes triggered by spreading seizures is the important goal of clinical neurology. Since apoptosis in one of processes occurring in epileptogenic focus and remote areas, treatment with such neuroprotective compounds as CsA and FK506 could minimise post-epileptic tissue deterioration.

In order to test this hypothesis, CsA or FK506 were administered to animals with pilocarpineinduced seizures when seizures were of intermediate intensity according to Racine's scale^{23,68}. Subsequently, the animals were observed for seizure progression. These experiments demonstrated that FK-506-treated animals showed lighter seizures than control animals and survived significantly longer. CsA administration also alleviated seizure intensity and prolonaed survival time after seizure initiation⁷⁵. Therefore, it can be assumed that each of the used compounds had a beneficial effect on the course of experimentally induced seizures, making them milder. It seems that the compounds under discussion can be efficient only when they are administered in the acute phase of seizures. Thus, they can presumably be considered to be primary neuroprotectants, since they counteract initial phenomena leading to neuronal death consequent to the spread of excitatory wave triggered probably by activation of voltage-gated Na⁺ and Ca²⁺ channels or glutamate receptors⁷⁶.

The above-described experiment does not unequivocally answer the question whether the studied compounds acted as primary neuroprotectants or by inhibiting the cascade leading to cell death, thereby contributing to secondary neuroprotection. The studies were carried out only for the first six hours after seizure initiation, thus, they should be linked with phenomena occurring in the early phase of epileptic activity, like excessive stimulation or primary neurodegenerative changes⁷⁷. Since the used compounds have neuroprotective activity, it can be assumed that they can save neurons in epileptogenic focus or its vicinity from death or degeneration⁷⁸. However, characteristics of those cells can be permanently changed, which may constitute a source of epileptic activity or susceptibility to seizures in the future. This hypothesis can be verified by further experiments with longer survival time after seizure initiation. If we envisage using compounds of similar type (calcineurin inhibitors) in clinical practice, we will also have to check whether they do not elicit undesirable effects in combination with currently used antiepileptic drugs.

6. Alternative forms of neuroprotective intervention: endogenous neuroprotection

It should be remembered that numerous complex processes occurring in damaged nervous tissue (degeneration, glial activity, regeneration) activate different tissue factors. Some of them can afford neuronal protection, particularly under pathological conditions. The presence of these factors or their absence can be decisive for death or survival of neurons at risk. Since these factors are produced by the nervous tissue itself, they are endogenous agents, therefore, their positive effect can be termed endogenous neuroprotection. It was discovered that moderate physical exercise can be one of situations inducing release of such endogenous neuroprotective factors.

As well as improving physical fitness in humans, moderate physical exercise improves psychical well-being, elevates mood, alleviates depression and decreases aggression⁷⁹. Motor training elicits also a number of changes in the nervous system, alters the levels of neurotransmitters: glutamate, aspartate and serine⁸⁰, and modifies expression of endogenous growth factors^{81,82}. Furthermore, exercise promotes brain vascularisation⁸³⁻⁸⁶.

Many experiments investigating electrical activity of the brain have proven that exercise can terminate non-physiological discharges but they return when training is over⁸⁷.

Setkowicz and Mazur⁸⁸ investigated whether regular long-term exercise of moderate intensity (45-day training on treadmill and swimming) changed susceptibility of adult rats to pilocarpine-induced epilepsy. The obtained results unequivocally confirmed that moderate exercise elicited a significant positive effect, since the course of experimentally induced seizures was milder in trained animals than in controls (Fig. 2).

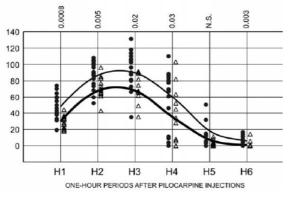


Fig. 2. Changes in the intensity of behavioural symptoms evoked by pilocarpine injections in trained and control, nontrained rats (thick and thin curves, respectively). Rating scores of the symptoms were summarised separately for each of six consecutive one-hour periods (H1-6) following pilocarpine injection. Triangles and circles represent the trained and control groups, respectively. Each decimal index shows statistical significance of difference between the two groups (Mann-Whitney test).

The neuroprotective effect of running may possibly be attributed to the formation of greater number of synapses⁸⁹, which results from the increased BDNF mRNA and BDNF expression in the hippocampus⁹⁰. BDNF was observed to inhibit excitotoxic changes in neuronal cultures⁹¹, and to reduce ischemia consequences *in vivo⁹²*. Therefore, it is believed that elevation of its level during exercise is accompanied by neuroprotective effect⁹³. It is also possible that exercise-induced rise in BDNF level triggers regulatory processes leading to the increase in neuropeptide Y level, which suppresses epileptic discharges⁹⁴. In addition, exercise enhances neurite growth factor (NGF) level. NGF can protect cells against hypoglycemia and excitotoxicity by reducing intracellular calcium excess⁹⁵. BDNF and NGF show also ability to boost activity of free radical scavengers⁹⁶, making the organism less endangered by their harmful effects. It is, thus, possible that, owning to the above-described properties, the NGF released during exercise abrogates pilocarpine-induced epileptic changes.

In other models of seizures, e.g. in the kindled seizures, trained animals were observed to require greater number of stimulations in order to elicit stage 5 seizures according to Racine's scale than non-trained animals⁹⁷, which corroborates positive effect of exercise.

Opposite effects were described by Ramsden et al.⁹⁸, who noted aggravation of neuronal damage after kainic acid-induced seizures in females subjected to physical exercise.

Advantages of endogenous over exogenous neuroprotection

Based on the above-described facts, it can be inferred that the application of substances commonly known to be neuroprotective after brain injury not necessarily has to have positive functional effects. Therefore, despite histological demonstration of distinctly increased neuronal survival in a damage area, this positive change may not be beneficial in terms of future susceptibility to epileptic attacks. The studies discussed in this review did not investigate such changes, so further experiments are necessary to clarify this issue. It is also required to examine the effects of vehicle components on development of epilepsy. Moreover, it is important to distinguish to what extent the neuroprotectant and the injury itself contribute to the observed changes. Therefore, CsA and FK506 effects should be examined also in the uninjured brain. Obviously, it is essential to take account of developmental changes occurring in the brain in the period when neuroprotective drugs are administered. Surprisingly, despite lack of these data, the analysed drugs have been used in posttransplational therapy in children for many years.

The administration of neuroprotective compounds in acute phase of seizures in animals brought positive effects and lowered seizure activity⁷⁵. However, long-term consequences of their use are uncertain, although their longlasting use in post-transplational therapy is known to be associated with the risk of different side effects^{99,100}.

Luckily, the stimulation of endogenous neuroprotection, like through exercise, seems propi-

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tious. Endogenous neuroprotection has no side effects accompanying the activation of various intracellular factors promoting neuronal survival. Furthermore, experiments refuted longstanding conviction that epileptic patients should avoid exercise. It may be expected that regular exercise of moderate intensity can counteract the increased susceptibility to seizures after injury. This hypothesis unquestionably requires experimental verification. Contemplating clinical use of neuroprotection free of side effects or invasive therapies, namely, endogenous neuroprotection, one should check whether the stimulation of intracellular repair systems is efficient in any experimental model of epilepsy. While attempting to apprise endogenous treatment strategies, one should take notice of the fact that all experiments presented here were carried out on males. Actually, epilepsy incidence in women and men is comparable, but experimental data indicate that seizure attacks in women are milder ¹⁰¹. Numerous researchers reported neuroprotective action of estrogen and progesterone^{102,103}. These hormones influence not only seizure intensity, but estrogen protects neurons from death by activation of *bcl* family genes or growth factors, and by inhibition of caspase-3, IL-1 and IL-6 activities¹⁰⁴. Furthermore, data from experiments on an animal model of traumatic brain injury indicate that progesterone significantly reduces neuronal damage in CA1 and CA2 and brain oedema¹⁰⁵.

Based on the above-presented facts, it can be assumed that the substances produced by the organism (neuropeptides, growth factors and even hormones) are more promising neuroprotectants for therapeutic prevention of generation and spread of epileptic stimulation in the brain than exogenous neuroprotective agents like FK506 or CsA. Researchers and clinicians need only to find a proper stimulation, assuring the adequate change in their level at critical time for seizure occurrence.

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References

- Arif H, Buchsbaum R, Weintraub D, Koyfman S, Salas-Humara C, Bazil CW, Resor SR, Hirsch LJ. Comparison and predictors of rash associated with 15 antiepileptic drugs. Neurology 2007;68:1701-9.
- (2) Loscher W.1997 Animal models of intractable epilepsy. Prog Neurobiol. 1997; 53: 239-58.
- (3) Turski WA, Cavalheiro EA, Bortolotto ZA, Mello LM, Schwarz M, Turski L. Seizures produced by pilocarpine in mice: a behavioral, electroencephalographic and morphological analysis.

Brain Res 1985;12:237-53.

- (4) Turski WA, Cavalheiro EA, Schwarz M, Czuczwar SJ, Kleinrok Z, Turski L. Limbic seizures produced by pilocarpine in rats: behavioural, electroencephalographic and neuropathological study. Behav Brain Res 1983; 9:315-35.
- (5) Turski L, Ikonomidou C, Turski WA, Bortolotto ZA, Cavalheiro EA. Review: cholinergic mechanisms and epileptogenesis. The seizures induced by pilocarpine: a novel experimental model of intractable epilepsy. Synapse1989;3:154-71.
- (6) Covolan L, Mello LE. Temporal profile of neuronal injury following pilocarpine or kainic acid-induced status epilepticus. Epilepsy Res 2000; 39 :133-52.
- (7) Kaminski RM, Blaszczak P, Dekundy A, Parada-Turska J, Calderazzo L, Cavalheiro EA, Turski WA. Lithium-methomyl induced seizures in rats: a new model of status epilepticus? Toxicol Appl Pharmacol 2007;219:122-7.
- (8) Cavalheiro EA, Leite JP, Bortolotto ZA, Turski WA, Ikonomidou C, Turski L. Long-term effects of pilocarpine in rats: structural damage of the brain triggers kindling and spontaneous recurrent seizures. Epilepsia1991;32:778-82.
- (9) Setkowicz Z., Kłak K. Janeczko K. Long-term changes in postnatal susceptibility to pilocarpine-induced seizures in rats exposed to gamma radiation at different stages of prenatal development. Epilepsia 2003;44:1267-1273.
- (10) Salazar AM, Jabbari B, Vance SC, Grafman J, Amin D, Dillon JD. Epilepsy after penetrating head injury. I. Clinical correlates: a report of the Vietnam Head Injury Study. Neurology 1985;35:1406-14.
- (11) Feeney DM, Walker AE. The prediction of posttraumatic epilepsy. A mathematical approach. Arch Neurol 1979;36:8-12.
- (12) Annegers JF, Coan SP. The risks of epilepsy after traumatic brain injury. Seizure 2000;9:453-7.
- (13) Frey LC. Epidemiology of posttraumatic epilepsy: a critical review. Epilepsia 2003;44 Suppl 10:11-7.
- (14) Gottesman RF, Komotar R, Hillis AE. Neurologic aspects of traumatic brain injury. Int Rev Psychiatry2003;15:302-9.
- (15) Hoffman SN, Salin PA, Prince DA. 1994 Chronic neocortical epileptogenesis in vitro. J Neurophysiol 1994; 71:1762-73.
- (16) D'Ambrosio R, Fairbanks JP, Fender JS, Born DE, Doyle DL, Miller JW.Post-traumatic epilepsy following fluid percussion injury in the rat. Brain 2004;127 :304-14.
- (17) Pitkanen A, McIntosh TK.2006 Animal models of post-traumatic epilepsy. 1: J Neurotrauma 2006; 23:241-61.
- (18) Dietrich AM, Bowman MJ, Ginn-Pease ME, Kosnik E, King DR. Pediatric head injuries: can clinical factors reliably predict an abnormality on computed tomography? Ann Emerg Med 1993;22:1535-40.
- (19) Janeczko K. The proliferative response of astrocytes to injury in neonatal rat brain. A combined immunocytochemical and autoradiographic study. Brain Res 1988;26:280-5.
- (20) Janeczko K. Spatiotemporal patterns of the astroglial proliferation in rat brain injured at

the postmitotic stage of postnatal development: a combined immunocytochemical and autoradiographic study. Brain Res1989;24:236-43.

- (21) Janeczko K. Co-expression of GFAP and vimentin in astrocytes proliferating in response to injury in the mouse cerebral hemisphere. A combined autoradiographic and double immunocytochemical study.Int J Dev Neurosci 1993;11:139-47.
- (22) Janeczko K. Age-dependent changes in the proliferative response of S-100 protein-positive glial cells to injury in the rat brain. Int J Dev Neurosci 1994;12:431-40.
- (23) Setkowicz Z. Janeczko K.Long-term changes in susceptibility to pilocarpine-induced status epilepticus following neocortical injures in the rat at different developmental stages. Epilepsy Res 2003;53:216-224.
- (24) Rice D, Barone S Jr. Critical periods of vulnerability for the developing nervous system: evidence from humans and animal models.Environ Health Perspect 2000;108 Suppl 3:511-33.
- (25) Penfield W. The physiology of epilepsy. Adv Neurol 1975;8:1-9.
- (26) Maxwell WL, Follows R, Ashhurst DE, Berry M. A The response of the cerebral hemisphere of the rat to injury. II. The neonatal rat.Philos Trans R Soc Lond B Biol Sci 1990; 26:501-13.
- (27) Acarin L, Gonzalez B, Castellano B. Neuronal, astroglial and microglial cytokine expression after an excitotoxic lesion in the immature rat brain. Eur J Neurosci 2001;12:3505-20.
- (28) Maxwell WL, Follows R, Ashhurst DE, Berry M.The response of the cerebral hemisphere of the rat to injury. I. The mature rat. Philos Trans R Soc Lond B Biol Sci 1990;26:479-500.
- (29) Tan AM, Zhang W, Levine JM. NG2: a component of the glial scar that inhibits axon growth. J Anat 2005;207:717-25.
- (30) D'Ambrosio R.The role of glial membrane ion channels in seizures and epileptogenesis. :Pharmacol Ther 2004;103:95-108.
- (31) Siesjo BK, Siesjo P.Mechanisms of secondary brain injury.Eur J Anaesthesiol 1996;13:247-68.
- (32) Setkowicz Z., Nowak B. Janeczko K.Neocortical injuries at different developmental stages determine different susceptibility to seizures induced in adulthood. Epilepsy Res 2006;68:255-263.
- (33) Ben-Ari Y. 1985 Limbic seizure and brain damage produced by kainic acid: mechanisms and relevance to human temporal lobe epilepsy. Neuroscience 1885 14:375-403.
- (34) Shiga Y, Onodera H, Matsuo Y, Kogure K.Cyclosporin A protects against ischemiareperfusion injury in the brain. Brain Res1992; 6;595:145-8.
- (35) Petcher TJ, Weber H, Rüegger A.Crystal and molecular structure of an iodo-derivative of the cyclic undecapeptide cyclosporin A. Helv Chim Acta 1976; 59:1480-9.
- (36) Ponticelli C. Cyclosporine: from renal transplantation to autoimmune diseases.Ann N Y Acad Sci 2005;1051:551-8.
- (37) Budde K, Bosmans JL, Sennesael J, Zeier M, Pisarski P, Schutz M, Fischer W, Neumayer HH, Glander P. Reduced-exposure cyclosporine is safe and efficacious in de novo renal trans-

plant recipients treated with enteric-coated mycophenolic acid and basiliximab. Clin Nephrol 2007;67:164-75.

- (38) Mihatsch MJ, Wolff K. Consensus conference on cyclosporin A for psoriasis February 1992 Br J Dermatol 1992;126:621-3.
- (39) Al-Daraji WI, Grant KR, Ryan K, Saxton A, Reynolds NJ. 2002 Localization of calcineurin/ NFAT in human skin and psoriasis and inhibition of calcineurin/NFAT activation in human keratinocytes by cyclosporin A. J Invest Dermatol. 118:779-88.
- (40) Yoshinoya S, Yamamoto K, Mitamura T, Aikawa T, Takeuchi A, Takahashi K, Miyamoto T. Successful treatment of rheumatoid arthritis with low-dose cyclosporine A. Transplant Proc 1988;20(3 Suppl 4):243-7.
- (41) Kino T, Hatanaka H, Hashimoto M, Nishiyama M, Goto T, Okuhara M, Kohsaka M, Aoki H, Imanaka H. FK-506, a novel immunosuppressant isolated from a Streptomyces. I. Fermentation, isolation, and physico-chemical and biological characteristics. J Antibiot (Tokyo) 1987;40:1249-55.
- (42) Ellis D. Clinical use of tacrolimus (FK-506) in infants and children with renal transplants. Pediatr Nephrol. 1995;9:487-94.
- (43) Schreiber SL, Crabtree GR. The mechanism of action of cyclosporin A and FK506. Immunol Today 1992;13:136-42.
- (44) Reynolds NJ, Al-Daraji WI.Calcineurin inhibitors and sirolimus: mechanisms of action and applications in dermatology. Clin Exp Dermatol 2002; 27 :555-61.
- (45) Galat A, Metcalfe SM.Peptidylproline cis/ trans isomerases. Prog Biophys Mol Biol 1995;63:67-118.
- (46) Yakel JL. Calcineurin regulation of synaptic function: from ion channels to transmitter release and gene transcription. Trends Pharmacol Sci 1995;18:124-34.
- (47) Cousin MA, Tan TC, Robinson PJ. 2001 Protein phosphorylation is required for endocytosis in nerve terminals: potential role for the dephosphins dynamin I and synaptojanin, but not AP180 or amphiphysin. J Neurochem 2001;76:105-16.
- (48) Dawson TM, Steiner JP, Dawson VL, Dinerman JL, Uhl GR, Snyder SH. Immunosuppressant FK506 enhances phosphorylation of nitric oxide synthase and protects against glutamate neurotoxicity. Proc Natl Acad Sci U S A 1993;1:9808-12.
- (49) Suzuki K, Sato M, Morishima Y, Nakanishi S. Neuronal depolarization controls brainderived neurotrophic factor-induced upregulation of NR2C NMDA receptor via calcineurin signaling. J Neurosci 2005;12:9535-43.
- (50) Rao A, Luo C, Hogan PG.Transcription factors of the NFAT family: regulation and function. Annu Rev Immunol 1997;15:707-47.
- (51) Clipstone NA, Crabtree GR.1992 Identification of calcineurin as a key signalling enzyme in T-lymphocyte activation. Nature 1992;25:695-7.
- (52) Rezzani R. Cyclosporine A and adverse effects on organs: histochemical studies.Prog Histochem Cytochem 2004;39:85-128.
- (53) Barton K, Muthusamy N, Chanyangam M,

Fischer C, Clendenin C, Leiden JM. 1996 Defective thymocyte proliferation and IL-2 production in transgenic mice expressing a dominant-negative form of CREB. Nature. 1996 379 :81-5.

- (54) Kruger M, Schwaninger M, Blume R, Oetjen E, Knepel W. Inhibition of CREB- and cAMP response element-mediated gene transcription by the immunosuppressive drugs cyclosporin A and FK506 in T cells.Naunyn Schmiedebergs Arch Pharmacol 1997;356:433-40.
- (55) Miyata K, Omori N, Uchino H, Yamaguchi T, Isshiki A, Shibasaki F. Involvement of the brainderived neurotrophic factor/TrkB pathway in neuroprotecive effect of cyclosporin A in forebrain ischemia. Neuroscience 2001;105:571-8.
- (56) Bito H, Deisseroth K, Tsien RW.CREB phosphorylation and dephosphorylation: a Ca(2+)- and stimulus duration-dependent switch for hippocampal gene expression. Cell 1996;27:1203-14.
- (57) Hu BR, Fux CM, Martone ME, Zivin JA, Ellisman MH.Persistent phosphorylation of cyclic AMP responsive element-binding protein and activating transcription factor-2 transcription factors following transient cerebral. Neuroscience 1999;89:437-52.
- (58) Matsuura K, Makino H, Ogawa N.Cyclosporin A attenuates the decrease in tyrosine hydroxylase immunoreactivity in nigrostriatal dopaminergic neurons and in striatal dopamine content in rats with intrastriatal injection of 6-hydroxydopamine.Exp Neurol1997;146:526-35.
- (59) Kuroda S, Janelidze S, Siesjo BK.The immunosuppressants cyclosporin A and FK506 equally ameliorate brain damage due to 30-min middle cerebral artery occlusion in hyperglycemic rats. Brain Res 1999;24:148-53.
- (60) Ibarra A, Correa D, Willms K, Merchant MT, Guizar-Sahagun G, Grijalva I, Madrazo I. 2003 Effects of cyclosporin-A on immune response, tissue protection and motor function of rats subjected to spinal cord injury. Brain Res 2001;25:165-78.
- (61) Gold BG, Katoh K, Storm-Dickerson T. The immunosuppressant FK506 increases the rate of axonal regeneration in rat sciatic nerve. J Neurosci 1995;15:7509-16.
- (62) Marmarou CR, Povlishock JT. Administration of the immunophilin ligand FK506 differentially attenuates neurofilament compaction and impaired axonal transport in injured axons following diffuse traumatic brain injury. Exp Neurol 2006;197:353-62.
- (63) Sharkey J, Butcher SP. Immunophilins mediate the neuroprotective effects of FK506 in focal cerebral ischaemia. Nature 1994; 22;371:336-9.
- (64) Butcher SP, Henshall DC, Teramura Y, Iwasaki K, Sharkey J.Neuroprotective actions of FK506 in experimental stroke: in vivo evidence against an antiexcitotoxic mechanism. J Neurosci 1997;15:6939-46.
- (65) Zawadzka M, Kaminska B. A novel mechanism of FK506-mediated neuroprotection: downregulation of cytokine expression in glial cells. Glia 2005;1:36-51.
- (66) Kaminska B, Gaweda-Walerych K, Zawadzka M. Molecular mechanisms of neuroprotective

action of immunosuppressants--facts and hypotheses.J Cell Mol Med. 2004 8 :45-58.

- (67) Yoshimoto T, Siesjo BK. Posttreatment with the immunosuppressant cyclosporin A in transient focal ischemia. Brain Res 1999;28:283-91.
- (68) Setkowicz Z, Ciarach M, Guzik R, Janeczko K. Different effects of neuroprotectants FK-506 and cyclosporin A on susceptibility to pilocarpine-induced seizures in rats with brain injured at different developmental stages. Epilepsy Res 2004;61:63-72.
- (69) Lee EJ, Lee SW, Choi HG, Kim CK.Bioavailability of cyclosporin A dispersed in sodium lauryl sulfate-dextrin based solid microspheres. Int J Pharm 2001;7:125-31.
- (70) Gelderblom H, Verweij J, Nooter K, Sparreboom A.Cremophor EL: the drawbacks and advantages of vehicle selection for drug formulation. Eur J Cancer 2001;37:1590-8.
- (71) Ran Y, Zhao L, Xu Q, Yalkowsky SH. Solubilization of cyclosporin A. AAPS PharmSciTech 2001;18:E2.
- (72) Strickley RG.Solubilizing excipients in oral and injectable formulations. Pharm Res 2004;21:201-30.
- (73) Fujikawa DG. The temporal evolution of neuronal damage from pilocarpine-induced status epilepticus. Brain Res1996;24:11-22.
- (74) Nevander G, Ingvar M, Auer R, Siesjo BK. Status epilepticus in well-oxygenated rats causes neuronal necrosis. Ann Neurol 1985;18:281-90.
- (75) Setkowicz Z. Ciarach M.Neuroprotectants FK-506 and cyclosporin A ameliorate the course of pilocarpine-induced seizures. Epilepsy Res 2007;73:151-155.
- (76) Meldrum BS. Concept of activity-induced cell death in epilepsy: historical and contemporary perspectives. Prog Brain Res 2002;135:3-11.
- (77) Fujikawa DG.Prolonged seizures and cellular injury: understanding the connection. 1: Epilepsy Behav 2005;7:S3-11.
- (78) Sanchez RM, Dai W, Levada RE, Lippman JJ, Jensen FE. AMPA/kainate receptor-mediated downregulation of GABAergic synaptic transmission by calcineurin after seizures in the developing rat brain. J Neurosci 2005; 30:3442-51.
- (79) Martinsen EW, Medhus A, Sandvik L.Effects of aerobic exercise on depression: a controlled study. Br Med J (Clin Res Ed) 1985;113;291:109.
- (80) Bland ST, Gonzale RA, Schallert T.Movementrelated glutamate levels in rat hippocampus, striatum, and sensorimotor cortex. 1: Neurosci Lett 1999;24:119-22.
- (81) Bortz WM, Angwin P, Mefford IN, Boarder MR, Noyce N, Barchas JD. Catecholamines, dopamine, and endorphin levels during extreme exercise. N Engl J Med 1981;20:466-7.
- (82) Gomez-Pinilla FV, Kesslak JP. Spatial learning and physical activity contribute to the induction of fibroblast growth factor: neural substrates for increased cognition associated with exercise. Neuroscience 1998;85:53-61.
- (83) Black JE, Isaacs KR, Anderso BJ, Alcantara AA, Greenough WT. Learning causes synaptogenesis, whereas motor activity causes angiogenesis, in cerebellar cortex of adult rats. Proc Natl Acad Sci U S A 1990;87:5568-72.

- (84) Isaacs KR, Anderson BJ, Alcantara AA, Black JE, Greenough WT. Exercise and the brain: angiogenesis in the adult rat cerebellum after vigorous physical activity and motor skill learning. J Cereb Blood Flow Metab 1992;12:110-9J.
- (85) Kleim JA, Cooper NR, VandenBerg PM. 2002. Exercise induces angiogenesis but does not alter movement representations within rat motor cortex. Brain Res 2002;26:1-6.
- (86) Swain RA, Harris AB, Wiener EC, Dutka MV, Morris HD, Theien BE, Konda S, Engberg K, Lauterbur PC, Greenough WT. Prolonged exercise induces angiogenesis and increases cerebral blood volume in primary motor cortex of the rat. Neuroscience 2003;117:1037-46.
- (87) Gotze W, Kubicki S, Munter M, Teichmann J. Effect of physical exercise on seizure threshold (investigated by electroencephalographic telemetry) Dis Nerv Syst 1967;28:664-7.
- (88) Setkowicz Z, Mazur A. Physical training decreases susceptibility to subsequent pilocarpine-induced seizures in the rat. Epilepsy Res 2006;71:142-148.
- (89) McAllister A.K, Katz LC, Lo DC. Neurotrophins and synaptic plasticity. Annu Rev Neurosci 1999;22:295-318.
- (90) Molteni R, Ying Z, Gomez-Pinilla F. Differential effects of acute and chronic exercise on plasticity-related genes in the rat hippocampus revealed by microarray. Eur J Neurosci 2002;16:1107-16.
- (91) Mattson MP, Lovell MA, Furukawa K, Markesber WR.Neurotrophic factors attenuate glutamate-induced accumulation of peroxides, elevation of intracellular Ca2+ concentration, and neurotoxicity and increase antioxidant enzyme activities in hippocampal neurons. J Neurochem 1995;165:1740-51.
- (92) Hefti F. Neurotrophic factor therapy for nervous system degenerative diseases. J Neurobiol 1994;25:1418-35.
- (93) Oliff HS, Berchtold NC, Isackson P, Cotman CW 1998 Exercise-induced regulation of brainderived neurotrophic factor (BDNF) transcripts in the rat hippocampus.Brain Res Mol Brain Res 1998;30:147-53.
- (94) Vezzani A, Ravizza T, Moneta D, Conti M, Borroni A, Rizzi M, Samanin R, Maj R. Brain-derived neurotrophic factor immunoreactivity in the limbic system of rats after acute seizures and during spontaneous convulsions: temporal evolution of changes as compared to neuropeptide Y.Neuroscience 1999;90:1445-61.
- (95) Cheng B, Mattso MP. NGF and bFGF protect rat hippocampal and human cortical neurons against hypoglycemic damage by stabilizing calcium homeostasis. Neuron 1991;7:1031-41.
- (96) Nistico G, Ciriolo MR, Fiskin K, Iannone M, De Martino A, Rotilio G. NGF restores decrease in catalase activity and increases superoxide dismutase and glutathione peroxidase activity in the brain of aged rats. Free Radic Biol Med 1992;12:177-81.
- (97) Arida RM, Fernandes MJ, Scorza FA, Preti SC, Cavalheiro EA. Physical training does not influence interictal LCMRglu in pilocarpinetreated rats with epilepsy.Physiol Behav 2003;79:789-94.
- (98) Ramsden M, Shin TM, Pike CJ. Androgens

modulate neuronal vulnerability to kainate lesion. Neuroscience 2003;122:573-8.

- (99) Wijdicks EF. Neurotoxicity of immunosuppressive drugs. Liver Transpl 2001;7:937-942.
- (100) Gijtenbeek JM, van den Bent MJ, Vecht CJ.Cyclosporine neurotoxicity: a review. J. Neurol 1999; 246,339-346.
- (101) Mejias-Aponte CA, Jimenez-Rivera CA, Segarra AC. 2002 Sex differences in models of temporal lobe epilepsy: role of testosterone. Brain Res 2002;19:210-8.
- (102) Green PS, Simpkins JW.Neuroprotective effects of estrogens: potential mechanisms of action. Int J Dev Neurosci 2000;18:347-58.
- (103) Roof RL, Hall ED. 2000 Gender differences in acute CNS trauma and stroke: neuroprotective effects of estrogen and progesterone. J Neurotrauma 2000;17:367-88.
- (104) Hoffman GE, Merchenthaler I, Zup SL.Endocrine 2006;29:217-31
- (105) Robertson CL, Puskar A, Hoffman GE, Murphy AZ, Saraswati M, Fiskum G 2006 Physiologic progesterone reduces mitochondrial dysfunction and hippocampal cell loss after traumatic brain injury in female rats. Exp Neurol 2006;197:235-43.